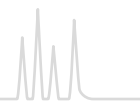




# Phase overview for special separations



Overview			
Separation / mechanism	Recommended column	Specification of the phase	Page
<b>Environmental analysis</b>			
Anion exchange chromatography of inorganic anions	NUCLEOGEL® Anion I	Strongly basic polymer-based anion exchanger	230
	NUCLEOSIL® Anion II	Strongly basic silica-based anion exchanger	
RP chromatography of PAHs	NUCLEODUR® C <sub>18</sub> PAH	NUCLEODUR® polymer-coated with C <sub>18</sub> groups USP L1	227
	NUCLEOSIL® 100-5 C <sub>18</sub> PAH	NUCLEOSIL® 100 polymer-coated with C <sub>18</sub> groups USP L1	229
<b>Enantiomer separation</b>			
Polar and $\pi$ - $\pi$ interactions	NUCLEOCEL DELTA	Silica-based modified cellulose phases USP L40	233
Formation of inclusion complexes	NUCLEODEX $\alpha$ -PM, $\beta$ -PM, $\gamma$ -PM and $\beta$ -OH	Silica-based permethylated and underivatized cyclodextrin phases USP L45	231
Enantioselective binding to chiral protein surface structures	RESOLVOSIL BSA-7	Silica-based protein phase (BSA)	234
Ligand exchange	NUCLEOSIL® CHIRAL-1	Covalently bonded amino acid – Cu(II) complexes USP L32	235
Charge-transfer, dipole-dipole interactions and others	NUCLEOSIL® CHIRAL-2	Silica-based brush type phases USP L36	236
	NUCLEOSIL® CHIRAL-3		
<b>Separation of biological macromolecules</b>			
Anion exchange chromatography of oligonucleotides and nucleic acids	NUCLEOGEN® DEAE	Silica-based DEAE anion exchanger	237
Anion exchange chromatography of peptides, large proteins and oligonucleotides	NUCLEOGEL® SAX	Polymer-based strongly basic anion exchanger USP L23	240
Cation exchange chromatography of proteins, peptides and carbohydrates	NUCLEOGEL® SCX	Polymer-based strong cation exchanger USP L22	240
	NUCLEOSIL® MPN	Monomerically bonded alkyl chains on silica USP L1 / USP L26	243
Reversed phase chromatography of proteins, peptides and oligonucleotides	NUCLEOSIL® PPN	PolymERICALLY bonded alkyl chains on silica USP L1	244
	NUCLEOGEL® RP 300	Polystyrene – divinylbenzene polymer USP L21	245
Reversed phase chromatography of small molecules	NUCLEOGEL® RP 100	Small pore macroporous PS-DVB polymer USP L21	245
<b>Food analysis · sugars</b>			
RP chromatography of mono- and oligosaccharides	NUCLEOSIL® Carbohydrate	Silica-based special amino phase USP L8	246
Separation of sugars, alcohols, org. acids based on ion exclusion, ion exchange, size exclusion, ligand exchange, NP and RP effects	NUCLEOGEL® SUGAR 810 H, Ca	Resins with sulfonic acid modification in different ionic forms H form USP L17 / Ca form L19 / Pb form L34 / Na form L58	247
Separation of sugars, alcohols, org. acids based on steric exclusion, ligand exchange and partition effects	NUCLEOGEL® SUGAR Ca, Na, Pb NUCLEOGEL® ION 300 OA		248
<b>Gel permeation chromatography (GPC)</b>			
Water-insoluble compounds	NUCLEOGEL® GPC	Polystyrene – divinylbenzene polymer	249



## NUCLEODUR® C<sub>18</sub> PAH special octadecyl phase for PAH analysis · USP L1

### Technical data

- Base material NUCLEODUR® silica, particle sizes 1.8 and 3 µm, pore size 110 Å; polymeric coating

### Recommended application

- Allows efficient gradient separation of the 16 PAHs according to EPA

### Analysis of 16 EPA PAHs with or without acetonitrile

MN Appl. Nos. 123820/123830

#### Separation with acetonitrile

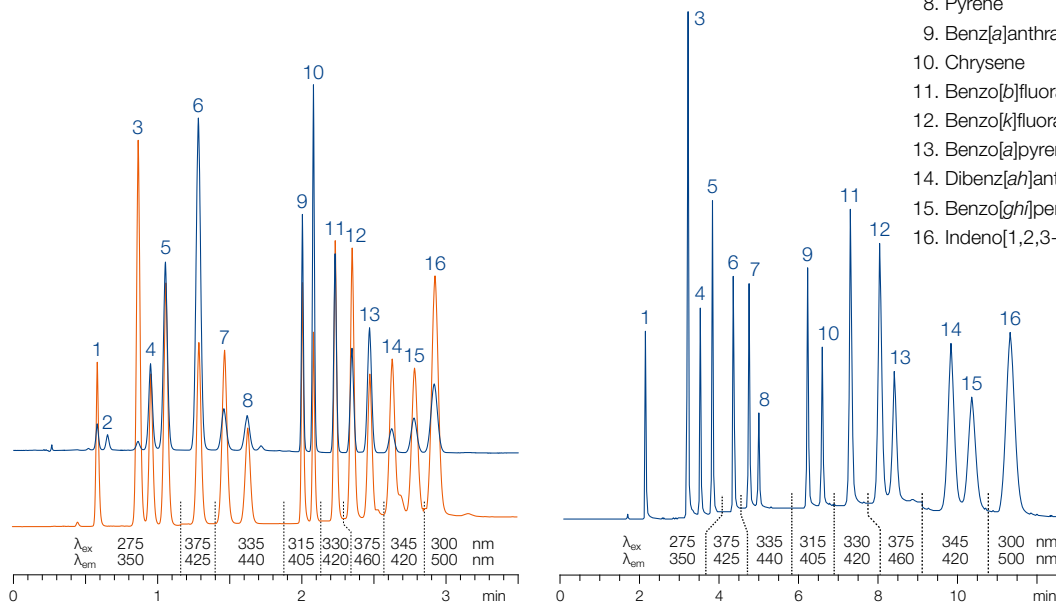
Column: 100 x 4 mm  
NUCLEODUR® C<sub>18</sub> PAH, 3 µm  
Eluent: A) methanol – water (80:20, v/v)  
B) acetonitrile 2–20% B in 1.2 min,  
20–100% B in 0.5 min, 100% B  
for 2.5 min, 100–2% B in 0.4 min  
Flow rate: 2.5 mL/min, temperature 35 °C  
Detection: UV, 254 nm  
fluorescence (see chromatogram)

#### Separation without acetonitrile

Column: 125 x 4 mm  
NUCLEODUR® C<sub>18</sub> PAH, 3 µm  
Eluent: A) water  
B) methanol 65–97% B in 6 min,  
97% B for 5 min, 97–65% B in  
0.5 min  
Flow rate: 2 mL/min, temperature 35 °C  
Detection: fluorescence (see chromatogram)

#### Peaks:



1. Naphthalene
2. Acenaphthylene (not detectable by fluorescence)
3. Acenaphthene
4. Fluorene
5. Phenanthrene
6. Anthracene
7. Fluoranthene
8. Pyrene
9. Benz[a]anthracene
10. Chrysene
11. Benzo[b]fluoranthene
12. Benzo[k]fluoranthene
13. Benzo[a]pyrene
14. Dibenzo[ah]anthracene
15. Benzo[ghi]perylene
16. Indeno[1,2,3-cd]pyrene



Detection of separated PAHs with UV (250–280 nm), diode array or fluorescence detection at different wavelengths for excitation and emission (acenaphthylene cannot be analyzed with fluorescence detection).

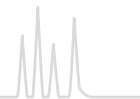
### Ordering information

Eluent in column acetonitrile – water (70:30, v/v)

ID	Length →					EC guard columns*
	100 mm	125 mm	150 mm	250 mm		
<b>NUCLEODUR® C<sub>18</sub> PAH, 1.8 µm</b> particle size 1.8 µm · UHPLC						
Analytical EC columns						
	2 mm	760773.20				761970.20
	3 mm	760773.30				761970.30
	4 mm	760773.40				761970.30
<b>NUCLEODUR® C<sub>18</sub> PAH, 3 µm</b> particle size 3 µm						
Analytical EC columns						
	3 mm	760783.30	760784.30	760785.30	760786.30	761971.30
	4 mm	760783.40	760784.40	760785.40	760786.40	761971.30

### Guard column system

Guard columns for EC columns with ID	2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC	4/2 (3)	4/3 (3)	4/3 (3)	718966



## Separation of 18 PAHs on NUCLEODUR® C<sub>18</sub> PAH

MN Appl. No. 123840

Column: 125 x 4 mm  
NUCLEODUR® C<sub>18</sub> PAH, 3 µm

Eluent: A) methanol – water  
(70:30, v/v); B) acetonitrile  
0–20 % B in 1.5 min,  
20–50 % B in 1.5 min,  
50–100 % B in 1.0 min,  
100 % B for 3 min,  
100–0 % B in 0.5 min

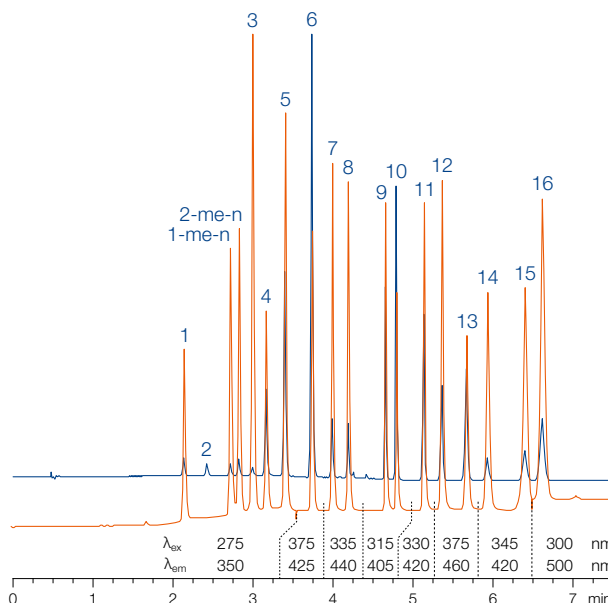
Flow rate: 1.5 mL/min

Temperature: 35 °C

Injection: UV: 1 µL,  
Fluorescence: 0.5 µL

Detection: UV, 254 nm  
fluorescence  
(see chromatogram)

Peaks:  
(concentrations 10 ng/µL per compound)  
1.–16. see page 227  
1-me-n: 1-methylnaphthalene  
2-me-n: 2-methylnaphthalene

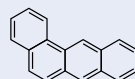


## Analysis of polycyclic aromatic hydrocarbons (PAHs) by HPLC

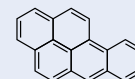
Polycyclic aromatic hydrocarbons (PAHs) are chemical compounds that consist of fused aromatic rings and do not contain heteroatoms or carry substituents. As a pollutant, they are of concern because some compounds have been identified as carcinogenic, mutagenic, and teratogenic. PAHs are natural components of coal or gas. They are delivered to our environment by pyrolysis (incomplete burning) of organic materials like coal, oil, fuel, wood, tobacco, ... and hence can be found globally. Today most PAHs accrue from anthropogenic processes – but also natural origins (forest fire) are possible. Regarding to past pollutions an important impact had production of coke and gas from black coal. Waste products (e.g., tar) from coking or gas plants are often origin of serious ground water pollutions.

Since a number of PAHs (e.g., benzo[a]pyrene, 3-methylcholanthrene and benzantracene) have been proven to be carcinogenic, control of the PAH content of food, water and soil is an important task for routine analysis. For choice and limiting values of the polycyclics we refer to the governmental regulations, which exist in many countries (e.g., EPA method 610 of the United States Environmental Protection Agency).

PAHs can be determined by different chromatographic techniques (TLC, GC, HPLC). Thus the 6 PAHs according to German drinking water specification (TVO) can, e.g., be analyzed by TLC (see German Standard DIN 38 409), while a much larger number of polycyclic aromatics can be determined by GC or HPLC.



Benzo[a]anthracen



Benzo[a]pyren

### HPLC columns for PAH analysis

For PAH analyses we have developed specially modified C<sub>18</sub> phases based on NUCLEODUR® and NUCLEOSIL® which allow efficient gradient separation of 16 PAHs according to EPA. Detection of the separated PAHs can be achieved by UV (250–280 nm), with diode array or with fluorescence detection at different wavelengths for excitation and emission. Acenaphthylene cannot be analyzed with fluorescence detection. For cost-effective routine PAH analysis we recommend applications using methanol instead of acetonitrile as eluent. For rapid analysis NUCLEODUR® C<sub>18</sub> PAH (3 µm) in short columns (100 mm) provides excellent results at high flow rates. Hereby separation of 16 PAHs according to EPA can be achieved in less than 3 min.

Tightened regulations require determination of 2 additional PAHs (1- and 2-methylnaphthalene) – so we developed highly efficient methods for 18 PAHs on the NUCLEODUR® C<sub>18</sub> PAH.



## NUCLEOSIL® 100-5 C<sub>18</sub> PAH special octadecyl phase for PAH analysis · USP L1

### Technical data

- Base material NUCLEOSIL® silica, particle size 5 µm, pore size 100 Å; polymeric coating
- Detection of the separated PAH with UV (250–280 nm), diode array or fluorescence detection at different wavelengths for excitation and emission (acenaphthylene cannot be analyzed with fluorescence detection)

### Recommended application

- Efficient gradient separation of the 16 PAHs according to EPA

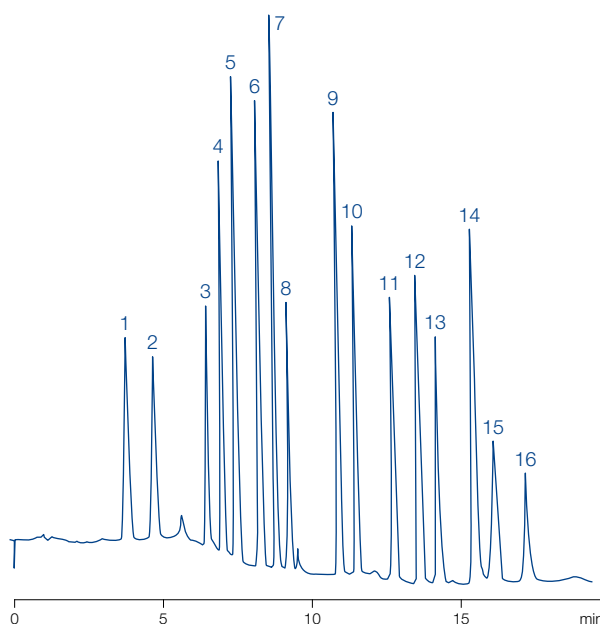
### Separation of the PAH standard according to EPA (REF 722393)

MN Appl. No. 115040

Column: 150 x 4 mm NUCLEOSIL® 100-5 C<sub>18</sub> PAH  
 Eluent: A) methanol – water (80:20)  
 B) acetonitrile – tetrahydrofuran (93:7)  
 0–100 % B in 10 min, 5 min 100 % B  
 Flow rate: 1 mL/min  
 Pressure: 140 bar  
 Temperature: 20 °C  
 Detection: UV, 260 nm


Peaks: (10 µg/mL each in acetonitrile)

- |                      |                            |
|----------------------|----------------------------|
| 1. Naphthalene       | 10. Chrysene               |
| 2. Acenaphthylene    | 11. Benzo[b]fluoranthene   |
| 3. Acenaphthene      | 12. Benzo[k]fluoranthene   |
| 4. Fluorene          | 13. Benzo[a]pyrene         |
| 5. Phenanthrene      | 14. Dibenz[ah]anthracene   |
| 6. Anthracene        | 15. Benzo[ghi]perylene     |
| 7. Fluoranthene      | 16. Indeno[1,2,3-cd]pyrene |
| 8. Pyrene            |                            |
| 9. Benz[a]anthracene |                            |



### Ordering information

Eluent in column acetonitrile – water 70:30

ID	Length →		
	150 mm	250 mm	EC guard columns*
<b>NUCLEOSIL® 100-5 C<sub>18</sub> PAH</b> particle size 5 µm, pore size 100 Å			
Analytical EC columns			
	2 mm	720117.20	721168.20
	3 mm	720923.30	721168.30
	4 mm	720923.40	721168.30
	4.6 mm	720117.46	721168.30

### PAH standard according to EPA for HPLC

Analytical EC columns

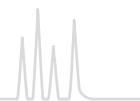
PAH standard for HPLC 16 PAH according to EPA method 610 in acetonitrile (1 mL) for composition see chromatogram above 722393

### Guard column system

Guard columns for EC columns with ID	2 mm	3 mm	4 mm	4.6 mm	Guard column holder
* Column Protection System (pack of)	EC 4/2 (3)	4/3 (3)	4/3 (3)	4/3 (3)	718966

EC columns in packs of 1, guard columns in packs of 3. For details of our column systems see page 250.

# This product contains harmful substances which must be specially labeled as hazardous. For detailed information please see SDS.



## Anion columns for analysis of inorganic anions

### NUCLEOGEL® Anion I

#### Technical data

- Strongly basic polymer-based anion exchanger, particle size 10 µm; pH stability 1–14
- Eluent in column 4 mmol/L salicylate buffer pH 7.8
- Contrary to the silica-based phase also suited for fluoride analysis

### NUCLEOSIL® Anion II

#### Technical data

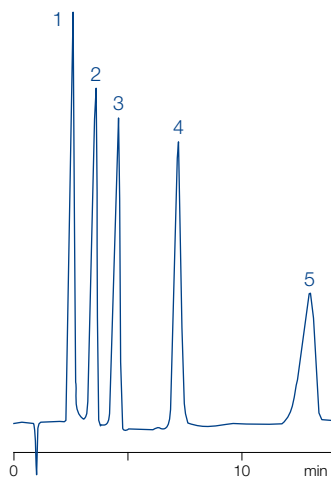
- Base material NUCLEOSIL® silica, particle size 10 µm, pore size 300 Å strongly basic anion exchanger, exchange capacity 50 µeq/g, pH stability 2–7.5
- Eluent in column 0.15 mol/L (NH<sub>4</sub>)<sub>2</sub>HPO<sub>4</sub> buffer pH 5.2 recommended buffer concentration for separation of inorganic anions: 2 mmol/L phthalate
- Preferred method of detection: conductivity or negative UV detection

#### Separation of an anion standard

MN Appl. No. 106440

Column: 250 x 4 mm NUCLEOSIL® Anion II  
 Eluent: 2 mmol/L potassium hydrogen phthalate, pH 5.7  
 Flow rate: 2 mL/min  
 Detection: UV, 280 nm

- Peaks:
1. H<sub>2</sub>PO<sub>4</sub><sup>-</sup>
  2. Cl<sup>-</sup>
  3. NO<sub>2</sub><sup>-</sup>
  4. NO<sub>3</sub><sup>-</sup>
  5. SO<sub>4</sub><sup>2-</sup>

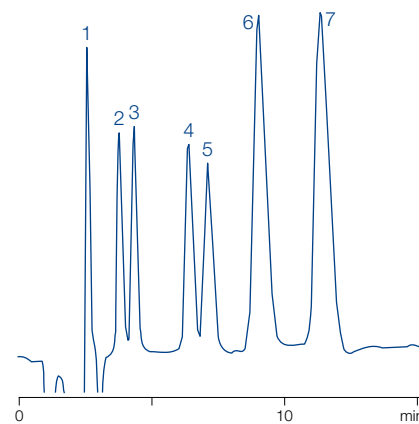


#### Separation of inorganic anions



MN Appl. No. 115050

Column: 120 x 4.6 mm NUCLEOGEL® Anion I  
 Eluent: 4 mmol/L salicylic acid – Tris pH 7.8  
 Flow rate: 1 mL/min  
 Detection: UV, 254 nm

- Peaks:
1. F<sup>-</sup>
  2. Cl<sup>-</sup>
  3. NO<sub>2</sub><sup>-</sup>
  4. Br<sup>-</sup>
  5. NO<sub>3</sub><sup>-</sup>
  6. PO<sub>4</sub><sup>3-</sup>
  7. SO<sub>4</sub><sup>2-</sup>



## Ordering information

ID	Length →		
	120 mm	250 mm	Guard columns*
<b>NUCLEOGEL® Anion I</b> eluent 4 mmol/L salicylate buffer pH 7.8			
Analytical Valco type columns			
 4.6 mm	719533		719543
<b>NUCLEOSIL® Anion II</b> eluent 0.15 mol/L (NH <sub>4</sub> ) <sub>2</sub> HPO <sub>4</sub> buffer pH 5.2			
Analytical EC columns			
 4 mm		720094.40	721169.30

\* NUCLEOGEL® Anion I Valco type guard columns cartridges are 21 x 4 mm, require guard column holder C, REF 719538, see page 250 (columns in packs of 1, guard columns in packs of 2)  
 NUCLEOSIL® Anion II guard columns are used with the Column Protection System (REF 718966, see page 251).



## NUCLEODEX columns enantiomer separation based on cyclodextrins

### NUCLEODEX $\beta$ -OH $\beta$ -cyclodextrin (R = H; n = 2) · USP L45

#### Technical data

- Base material NUCLEOSIL® silica, particle size 5  $\mu\text{m}$ , pore size 100 Å modified cyclodextrins as chiral selectors
- Separation based on hydrogen bonds and dipole interactions between functional groups of the analyte and hydroxyl groups of the cyclodextrin
- Examples for successful enantiomer separations: chlorthalidone and other compounds, which require free hydroxyl groups for enantioselective interactions
- Eluent in column  $\text{CH}_3\text{OH}$  – 0.1 % TEAA pH 4 (55:45)

### NUCLEODEX $\alpha$ -PM permethylated $\alpha$ -cyclodextrin (R = $\text{CH}_3$ ; n = 1)

#### Technical data

- Base material NUCLEOSIL® silica, particle size 5  $\mu\text{m}$ , pore size 100 Å modified cyclodextrins as chiral selectors
- Examples for successful enantiomer separations: mecoprop and dichlorprop as free carboxylic acids, trans-stilbene oxide, styrene oxide
- Eluent in column  $\text{CH}_3\text{OH}$  – 50 mmol/L phosphate pH 3 (70:30)

### NUCLEODEX $\beta$ -PM permethylated $\beta$ -cyclodextrin (R = $\text{CH}_3$ ; n = 2) · USP L45

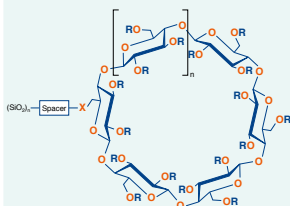
#### Technical data

- Base material NUCLEOSIL® silica, particle size 5  $\mu\text{m}$ , pore size 100 Å modified cyclodextrins as chiral selectors
- Examples for successful enantiomer separations: mephobarbital (prominal), pesticide derivatives mecoprop methyl and dichlorprop methyl
- Eluent in column  $\text{CH}_3\text{OH}$  – 0.1 % TEAA pH 4 (65:35)

### NUCLEODEX $\gamma$ -PM permethylated $\gamma$ -cyclodextrin (R = $\text{CH}_3$ ; n = 3)

#### Technical data

- Base material NUCLEOSIL® silica, particle size 5  $\mu\text{m}$ , pore size 100 Å modified cyclodextrins as chiral selectors
- Examples for successful enantiomer separations: steroids or other larger molecules
- Eluent in column  $\text{CH}_3\text{OH}$  – 0.1 % TEAA pH 4 (55:45)

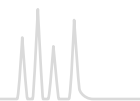


#### Recommended application

- NUCLEODEX phases are especially suited for the control of optical purity, but also for semipreparative separations and for the analysis of positional and cis-trans isomers.
- For numerous separations on NUCLEODEX phases please visit our website: [www.mn-net.com/apps](http://www.mn-net.com/apps)



# HPLC columns for enantiomer separations

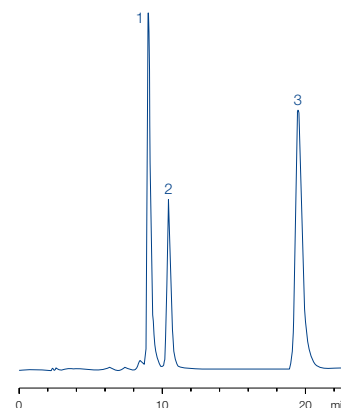


## Separation of the positional isomers of nitroaniline

MN Appl. No. 101420

Column: 200 x 4 mm NUCLEODEX β-OH  
 Eluent: methanol – 0.1 % triethylammonium acetate pH 4.0 (50:50, v/v)  
 Flow rate: 0.7 mL/min  
 Pressure: 180 bar  
 Detection: UV, 254 nm  
 Injection: 1 μL

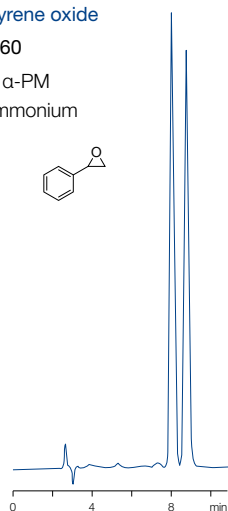
- Peaks:
1. *m*-Nitroaniline
  2. *o*-Nitroaniline
  3. *p*-Nitroaniline



## Enantiomer separation of styrene oxide

MN Appl. No. 106160

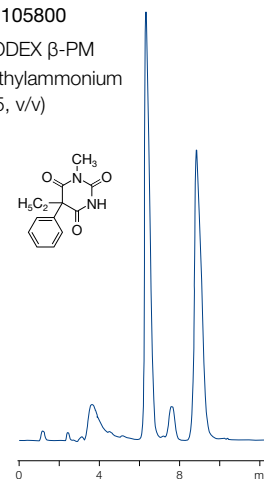
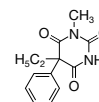
Column: 200 x 4 mm NUCLEODEX α-PM  
 Eluent: methanol – 0.1 % triethylammonium acetate pH 4.0 (60:40, v/v)  
 Flow rate: 0.7 mL/min  
 Pressure: 160 bar  
 Detection: UV, 230 nm  
 Injection: 2 μL



## Enantiomer separation of mephobarbital

MN Appl. No. 105800

Column: 200 x 4 mm NUCLEODEX β-PM  
 Eluent: methanol – 0.1 % triethylammonium acetate pH 4.0 (55:45, v/v)  
 Flow rate: 0.7 mL/min  
 Pressure: 180 bar  
 Detection: UV, 254 nm  
 Injection: 1 μL



## Ordering information

ID	Length → 200 mm	EC guard columns*
<b>NUCLEODEX β-OH</b> eluent methanol – 0.1 % TEAA pH 4 (55:45)		
Analytical EC columns		
4 mm	720124.40	721171.30
<b>NUCLEODEX α-PM</b> eluent methanol – 50 mmol/L phosphate pH 3 (70:30)		
Analytical EC columns		
4 mm	720127.40	721469.30
<b>NUCLEODEX β-PM</b> eluent methanol – 0.1 % TEAA pH 4 (65:35)		
Analytical EC columns		
4 mm	720125.40	721176.30
<b>NUCLEODEX γ-PM</b> eluent methanol – 0.1 % TEAA pH 4 (55:45)		
Analytical EC columns		
4 mm	720752.40	721178.30

## NUCLEODEX CC screening kit

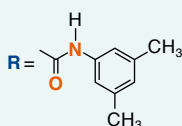
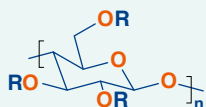
contains one CC 30/4 each with NUCLEODEX β-OH, α-PM, β-PM and γ-PM as well as one CC column holder 30 mm

721920

\* EC 4/3 guard columns for EC columns with 4 mm ID require the Column Protection System guard column holder (REF 718966, see page 251). Columns and guard columns in packs of 1.



## NUCLEOCEL DELTA enantiomer separation based on a cellulose derivative · USP L40



### Technical data

- Base material silica, chiral selector cellulose tris-(3,5-dimethylphenylcarbamate)
- High resolution type (S) with 5 µm particle size, allows use of shorter columns (150 mm) for faster separations, pressure stability up to ~150 bar (2000 psi), pH stability 1–9

NUCLEOCEL DELTA for normal phase applications: eluent in column *n*-heptane – 2-propanol (90:10, v/v) typical eluents are heptane – propanol mixtures

NUCLEOCEL DELTA-RP for reversed phase applications: eluent in column acetonitrile – water (40:60, v/v) designed for use either in polar organic mode or with eluents containing high concentrations of chaotropic salts such as perchlorate

### Recommended application

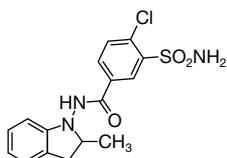
- Pharmaceutically active compounds, chiral pollutants (e.g., herbicides, PCB), chiral compounds in food (dyes, preservatives), chiral catalysts and bioorganic compounds

Similar phases: Chiralcel® OD, Kromasil® CelluCoat™, Eurocel® 01, Lux™ Cellulose-1

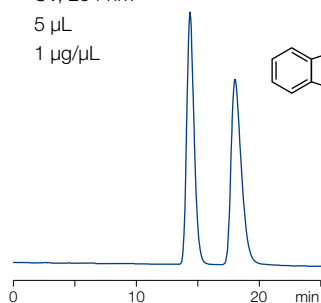
### Enantiomer separation of indapamide

MN Appl. No. 121230

Column: 250 x 4,6 mm NUCLEOCEL DELTA-RP S  
 Eluent: acetonitrile – water (40:60, v/v)  
 Flow rate: 0.5 mL/min  
 Temperature: 40 °C  
 Detection: UV, 254 nm  
 Injection: 5 µL  
 Concentration: 1 µg/µL



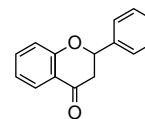
$\alpha = 1.3$   
 $R_s = 2.6$



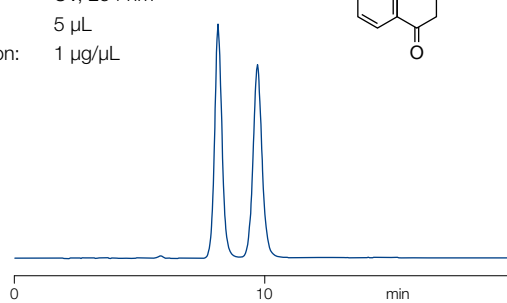
### Enantiomer separation of flavanone

MN Appl. No. 121260



Column: 250 x 4,6 mm NUCLEOCEL DELTA S  
 Eluent: *n*-heptane – 2-propanol (90:10, v/v)  
 Flow rate: 1 mL/min  
 Temperature: 25 °C  
 Detection: UV, 254 nm  
 Injection: 5 µL  
 Concentration: 1 µg/µL



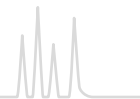
$\alpha = 1.29$   
 $R_s = 2.6$



### Ordering information

ID	Length →	250 mm		EC guard columns*
		150 mm	250 mm	
<b>NUCLEOCEL DELTA S, 5 µm</b> eluent <i>n</i> -heptane – 2-propanol (90:10, v/v)				
Analytical EC columns				
 4.6 mm			720445.46	721185.30
<b>NUCLEOCEL DELTA-RP S, 5 µm</b> eluent acetonitrile – water (40:60, v/v)				
Analytical EC columns				
 4.6 mm		720451.46	720450.46	721186.30

\* EC 4/3 guard column cartridges are used for EC columns of 4.6 mm ID with the Column Protection System guard column holder (REF 718966, see page 251). Columns and guard columns in packs of 1.



## RESOLVOSIL BSA-7 protein phase for enantiomer separation · USP L75

### Technical data

- Base material NUCLEOSIL® silica, particle size 7 µm, pore size 300 Å chiral selector bovine serum albumin (BSA)
- Separation based on selective interaction of proteins with low molecular compounds, i.e. principles of bioaffinity, including hydrophobic interactions (similar to a true reversed phase), interactions of polar groups and steric effects

### Recommended application

- Amino acid derivatives, aromatic amino acids, aromatic sulfoxides, barbiturates, benzodiazepinones, benzoin and benzoin derivatives, β-blockers, coumarin derivatives, and for monitoring stereoselective microbial and enzymatic conversions

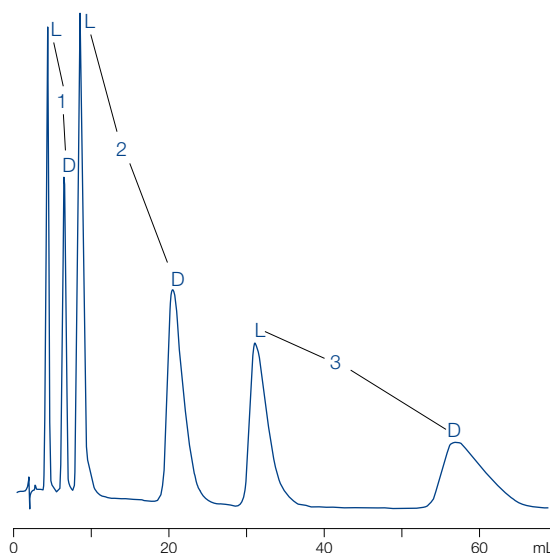
### Enantiomer separation of *N*-benzoyl-*D,L*-amino acids

MN Appl. No. 105450

S. Allenmark et al. in "Affinity chromatography and biological recognition" (I. Chaiken, M. Wilchek, and I. Parikh. Eds.), Academic Press, New York, 1983, 259–260

Column: 150 x 4 mm RESOLVOSIL BSA-7  
 Eluent: 50 mmol/L phosphate buffer pH 6.5  
 + 1 % 1-propanol  
 Flow rate: 0.70 mL/min  
 Detection: UV, 225 nm

- Peaks:
1. Serine
  2. Alanine
  3. Phenylalanine



### Ordering information

Eluent in column 0.1 mol/L phosphate buffer pH 7.5, 2 % 1-propanol

ID	Length → 150 mm	EC guard columns*
----	--------------------	-------------------

### RESOLVOSIL BSA-7

Analytical EC columns



4 mm

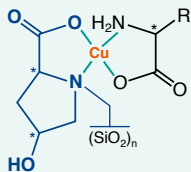
720046.40

721402.30

\* EC 4/3 guard columns for EC columns with 4 mm ID require the Column Protection System guard column holder (REF 718966, see page 251). Columns and guard columns in packs of 1.



## NUCLEOSIL® CHIRAL-1 enantiomer separation based on ligand exchange · USP L32



### 🔧 Technical data

- Base material NUCLEOSIL® silica, particle size 5 µm, pore size 120 Å chiral selector L-hydroxyproline – Cu<sup>2+</sup> complexes
- Principal interaction mode:
  - formation of ternary mixed-ligand complexes with Cu(II) ions; differences in the stability of the diastereomeric complexes cause chromatographic separation

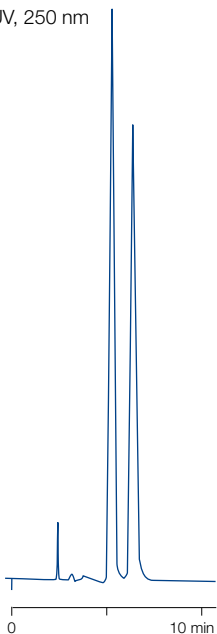
### ✓ Recommended application

- Enantiomers with two polar functional groups with the correct spacing such as α-amino acids, α-hydroxycarboxylic acids (e.g., lactic acid), *N*-alkyl-α-amino acids etc.

### *D,L*-alanine enantiomers

MN Appl. No. 105410

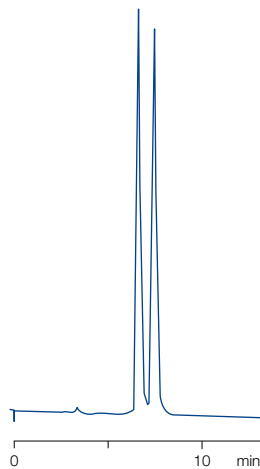
Column: 250 x 4 mm NUCLEOSIL® CHIRAL-1  
 Eluent: 0.5 mmol/L CuSO<sub>4</sub>  
 Flow rate: 1 mL/min  
 Pressure: 60 bar  
 Temperature: 60 °C  
 Detection: UV, 250 nm



### *D,L*-threonine enantiomers

MN Appl. No. 105410

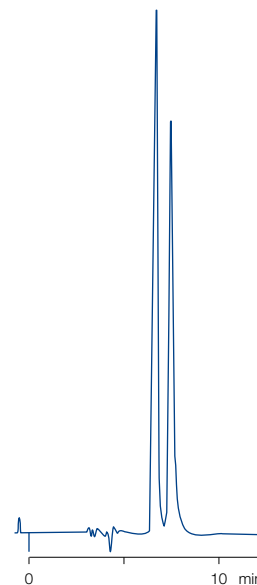
Column: 250 x 4 mm NUCLEOSIL® CHIRAL-1  
 Eluent: 0.25 mmol/L CuSO<sub>4</sub>  
 Flow rate: 0.8 mL/min  
 Pressure: 65 bar  
 Temperature: 60 °C  
 Detection: UV, 240 nm



### Lactic acid enantiomers

MN Appl. No. 105560

Column: 250 x 4 mm NUCLEOSIL® CHIRAL-1  
 Eluent: 0.5 mmol/L CuSO<sub>4</sub>  
 Flow rate: 0.8 mL/min  
 Temperature: 60 °C  
 Detection: UV, 240 nm  
 Injection: 1 µL



### Ordering information

Eluent in column 0.5 mmol/L copper sulfate solution

ID

Length →

250 mm

EC guard columns\*

### NUCLEOSIL® CHIRAL-1

Analytical EC columns

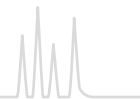


4 mm

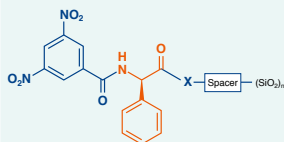
720081.40

721188.30

\* EC 4/3 guard columns for EC columns with 4 mm ID require the Column Protection System guard column holder (REF 718966, see page 251). Columns and guard columns in packs of 1.



## NUCLEOSIL® CHIRAL-2 · CHIRAL-3 enantiomer separation in organic eluent systems · USP L36



### Technical data

- Base material NUCLEOSIL® silica, particle size 5 µm, pore size 100 Å chiral selector for NUCLEOSIL® CHIRAL-2 is *N*-(3,5-dinitrobenzoyl)-*D*-phenylglycine, for CHIRAL-3 the optical antipode is used, “brush type” phases
- Principle interaction modes: charge-transfer interactions, hydrogen bonds, dipole-dipole interactions and steric effects

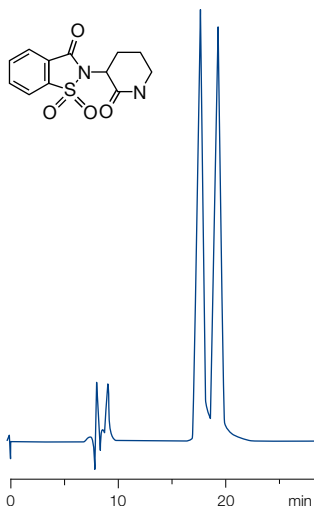
### Recommended application

- analysis of stereoisomers such as separation of enantiomers and diastereomers, control of optical purity of plant protectives (pesticides, e.g., propionic acid derived herbicides) pharmaceuticals etc. and for product control in chiral organic syntheses
- For control of optical purity of a substance, the columns NUCLEOSIL® CHIRAL-2 and NUCLEOSIL® CHIRAL-3 allow to select conditions such that the minor enantiomer, present as an impurity, is eluted before the main peak. Overlapping peaks are avoided. This makes an exact quantification of the impurity much easier.

### Enantiomer separation of *D,L*-supidimide

MN Appl. No. 105690

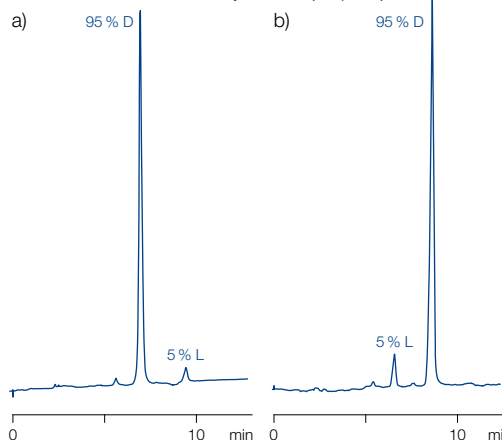
Column: 250 x 4 mm NUCLEOSIL® CHIRAL-2  
 Eluent: tetrahydrofuran – *n*-heptane (10:3, v/v)  
 Flow rate: 1.0 mL/min  
 Detection: UV, 220 nm



### Control of optical purity of mecoprop methyl

MN Appl. No. 111360

Columns: a) 250 x 4 mm NUCLEOSIL® CHIRAL-2  
 b) 250 x 4 mm NUCLEOSIL® CHIRAL-3  
 Eluent: *n*-heptane – 2-propanol – TFA (100:0.05:0.05, v/v/v)  
 Flow rate: 1 mL/min, ambient temperature  
 Detection: UV, 230 nm, Injection 1 µL (sample with 90 % ee)



### Ordering information

Eluent in column *n*-heptane – 2-propanol – TFAA (100:0.05:0.05, v/v/v)

ID	Length → 250 mm	EC guard columns*
<b>NUCLEOSIL® CHIRAL-2</b>		
Analytical EC columns		
4 mm	720088.40	721190.30
<b>NUCLEOSIL® CHIRAL-3</b>		
Analytical EC columns		
4 mm	720350.40	721190.30

Guard columns for NUCLEOSIL® CHIRAL-2 and CHIRAL-3 are identical.

\* EC 4/3 guard columns for EC columns with 4 mm ID require the Column Protection System guard column holder (REF 718966, see page 251). EC columns and EC guard columns in packs of 1.



## NUCLEOGEN® columns anion exchange chromatography of nucleic acids

### NUCLEOGEN® 60-7 DEAE pore size 60 Å

#### Technical data

- Base material silica, particle size 7 µm; DEAE anion exchanger
- For the separation of oligonucleotides up to chain lengths of 40 bases with recoveries > 95 % capacity 200 A<sub>260</sub>/mL (~ 300 A<sub>260</sub> for a 125 x 4 mm ID column, 1875 A<sub>260</sub> for a 125 x 10 mm ID column)
- Preparative separations possible when using higher flow rates and longer gradient times

### NUCLEOGEN® 500-7 DEAE pore size 500 Å

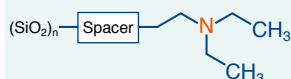
#### Technical data

- Base material silica, particle size 7 µm; DEAE anion exchanger
- For the separation of tRNA, 5S RNA, viroids and messenger RNA in the intermediate molecular weight range (25–1 000 kDa) with recoveries > 95 %
- Capacity 730 A<sub>260</sub> for a 125 x 6 mm ID column, 1940 A<sub>260</sub> for a 125 x 10 mm ID column

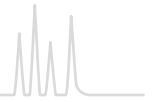
### NUCLEOGEN® 4000-7 DEAE pore size 4000 Å

#### Technical data

- Base material silica, particle size 7 µm; DEAE anion exchanger
- For the separation of plasmids, DNA restriction fragments, ribosomal RNA, messenger RNA and viral RNA, i.e. very high molecular weight nucleic acids (e.g., 1–50 MDa)
- Capacity 120 A<sub>260</sub> for a 125 x 6 mm ID column, 350 A<sub>260</sub> for a 125 x 10 mm ID column



For more separations of deoxyoligonucleotides, plasmids and DNA restriction fragments visit our website [www.mn-net.com/apps](http://www.mn-net.com/apps)



## Separation of plasmid pBR 322

MN Appl. No. 107480

M. Colpan, D. Riesner, private communication

A) isolation of plasmid DNA from a crude cell lysate

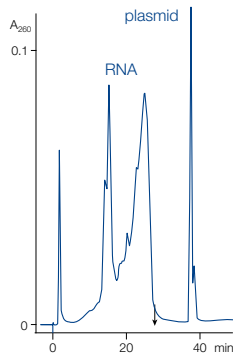
Sample: 5 µg plasmid pBR 322 containing cleared lysate from *E. coli*

Column: 125 x 6 mm NUCLEOGEN® 4000-7 DEAE

Eluent: A) 20 mmol/L K phosphate buffer pH 6.9; 5 mol/L urea  
B) eluent A + 1.5 mol/L KCl  
20–100 % B in 50 min;  
arrow = ionic strength of 850 mmol/L

Flow rate: 1.0 mL/min, 70 bar, ambient temperature

Detection: UV, 260 nm



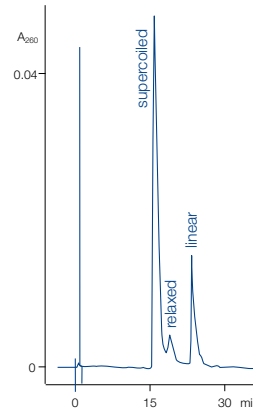
B) separation of supercoiled plasmid from relaxed and linear forms

Sample: plasmid pBR 322, supercoiled, relaxed and linear

Column: 125 x 6 mm NUCLEOGEN® 4000-7 DEAE

Eluent: A) 20 mmol/L K phosphate buffer pH 6.8; 6 mol/L urea  
B) eluent A + 2 mol/L KCl  
42–100 % B in 230 min

Flow rate: 1.5 mL/min, 45 bar, ambient temperature



## Separation of oligo(rA)<sub>n</sub>

MN Appl. No. 115180

Column: 125 x 4 mm NUCLEOGEN® 60-7 DEAE

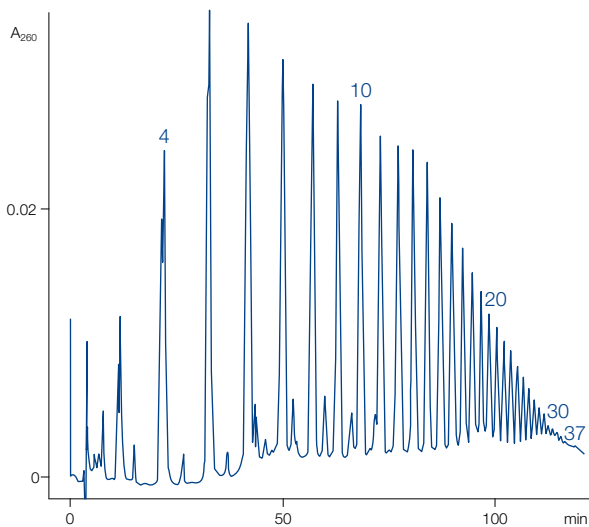
Eluent: A) 20 mmol/L phosphate buffer, pH 5.5, 5 mol/L urea  
B) buffer A + 1 mol/L KCl  
0–100 % B in 200 min

Flow rate: 2 mL/min

Pressure: 110 bar

Temperature: ambient

Detection: UV, 260 nm



## Preparative separation of a crude RNA extract of viroid (PSTV) infected tomato plants

MN Appl. No. 107490

D. Riesner, BioEngineering 1 (1988) 42–48

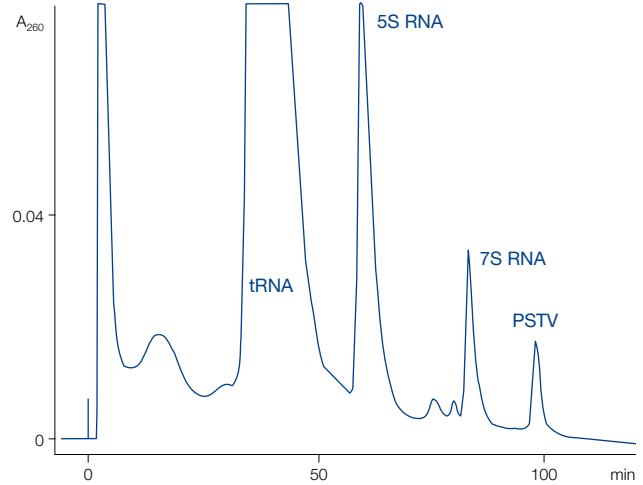
Column: 125 x 6 mm NUCLEOGEN® 500-7 DEAE

Eluent: A) 250 mmol/L KCl, 20 mmol/L phosphate buffer, pH 6.6, 5 mol/L urea  
B) 1 mol/L KCl, 20 mmol/L phosphate buffer, pH 6.6, 5 mol/L urea  
0–50 % B in 120 min, 50–100 % B in 250 min

Flow rate: 3 mL/min

Pressure: 40 bar, ambient temperature

Detection: 260 nm











# HPLC columns for biochemical separations

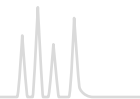


## Ordering information

Eluent in column methanol

ID	Length → 125 mm	Guard columns*
<b>NUCLEOGEN® 60-7 DEAE</b> particle size 7 µm, pore size 60 Å		
Analytical EC columns		
 4 mm	736596.40	736400.40
Preparative VarioPrep columns		
 10 mm	736597.100	736400.40
<b>NUCLEOGEN® 500-7 DEAE</b> particle size 7 µm, pore size 500 Å		
Analytical Valco type columns		
 6 mm	736598	736400.40
Preparative VarioPrep columns		
 10 mm	736599.100	736400.40
<b>NUCLEOGEN® 4000-7 DEAE</b> particle size 7 µm, pore size 4000 Å		
Analytical Valco type columns		
 6 mm	736601	736400.40
Preparative VarioPrep columns		
 10 mm	736602.100	736400.40

\* NUCLEOGEN® guard columns are 30 mm long and require the CC column holder 30 mm (REF 721823).  
Columns in packs of 1, guard columns in packs of 2.



## NUCLEOGEL® SAX anion exchange of biological macromolecules · USP L23

### Technical data

- Polymer-based strongly basic anion exchanger  $-N^+(CH_3)_3$ , gel matrix quaternized PEI; particle size 8  $\mu\text{m}$ , pore size 1000  $\text{\AA}$
- pH working range 1–13, max. working pressure 200 bar

### Recommended application

- Purification of peptides, large proteins and oligonucleotides, high capacity for proteins even at pH 10

## NUCLEOGEL® SCX cation exchange of biological macromolecules · USP L22

### Technical data

- Polymer-based strongly acidic cation exchanger  $-SO_3^-$ , hydrophilic gel matrix; particle size 8  $\mu\text{m}$ , pore size 1000  $\text{\AA}$
- pH working range 1–13, max. working pressure 200 bar

### Recommended application

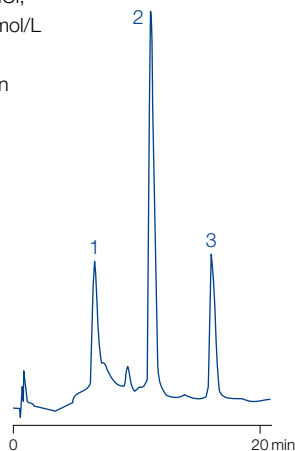
- Proteins, peptides and carbohydrates with high isoelectric point

### Separation of hen's egg white

MN Appl. No. 115200

Sample: frozen egg white was thawed, filtered and diluted 1:8 with eluent A  
 Column: 50 x 4.6 mm NUCLEOGEL® SAX 1000-8  
 Eluent: A) 0.01 mol/L Tris-HCl, pH 7.5; B) A + 0.5 mol/L NaAc, pH 7.5; 0–100 % B in 20 min  
 Flow rate: 1 mL/min  
 Inj. volumen: 50  $\mu\text{L}$   
 Detection: UV, 280 nm

- Peaks:
1. Conalbumin
  2. Ovalbumin
  3. not identified

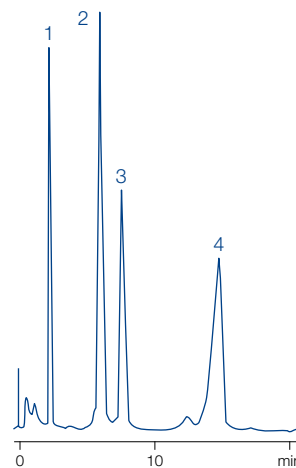


### Separation of protein standards

MN Appl. No. 108261

Column: 50 x 4.6 mm NUCLEOGEL® SCX 1000-8  
 Eluent: A) 0.02 mol/L  $\text{KH}_2\text{PO}_4$ , pH 6.0  
 B) A + 0,5 mol/L NaCl, pH 6.0  
 0–100 % B in 20 min  
 Flow rate: 1 mL/min  
 Detection: UV, 280 nm

- Peaks:
1. Myoglobin
  2.  $\alpha$ -Chymotrypsinogen A
  3. Cytochrome C
  4. Lysozyme



## Ordering information

Eluent in column 0.1 mol/L  $\text{Na}_2\text{SO}_4$  + 0.2 %  $\text{NaN}_3$

ID

Length →  
50 mm

Guard columns\*

### NUCLEOGEL® SAX pore size 1000 $\text{\AA}$

Analytical Valco type columns



4.6 mm

719469

719600

### NUCLEOGEL® SCX pore size 1000 $\text{\AA}$

Analytical Valco type columns



4.6 mm

719475

719540

\* NUCLEOGEL® SAX and SCX Valco type guard columns measure 5 x 3 mm and require the guard column holder B, REF 719539 (see page 250)  
 Columns in packs of 1, guard columns in packs of 2.



## NUCLEODUR® 300 C<sub>18</sub> ec · C<sub>4</sub> ec wide pore silica for biochromatography · USP L1 (C<sub>18</sub>) · USP L26 (C<sub>4</sub>)

### ★ Key feature

- Reliable wide pore RP phases for daily routine analysis
- Medium density octadecyl or butyl modification with exhaustive endcapping
- Ideal phases for separation of biomolecules

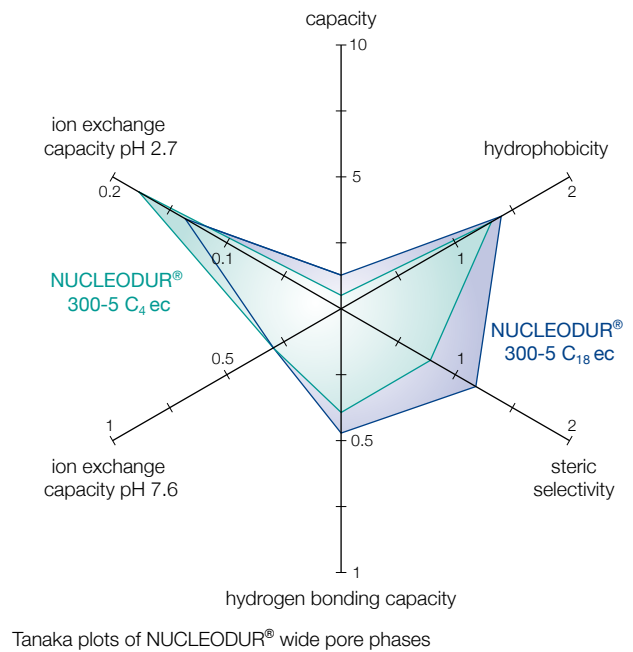
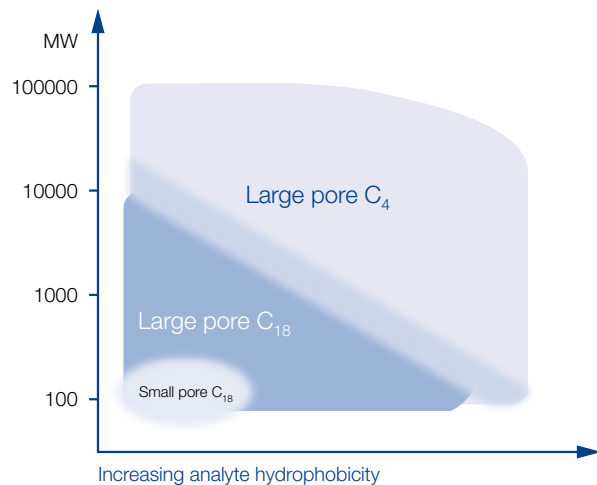
### 🔧 Technical data

- Pore size 300 Å; particle size 5 µm, carbon content 4 % for C<sub>18</sub>, 2.5 % for C<sub>4</sub>; pH stability 1–9; high reproducibility from lot to lot

### ✓ Recommended application

- Biological macromolecules like proteins or peptides

### Column selection by analyte characteristics

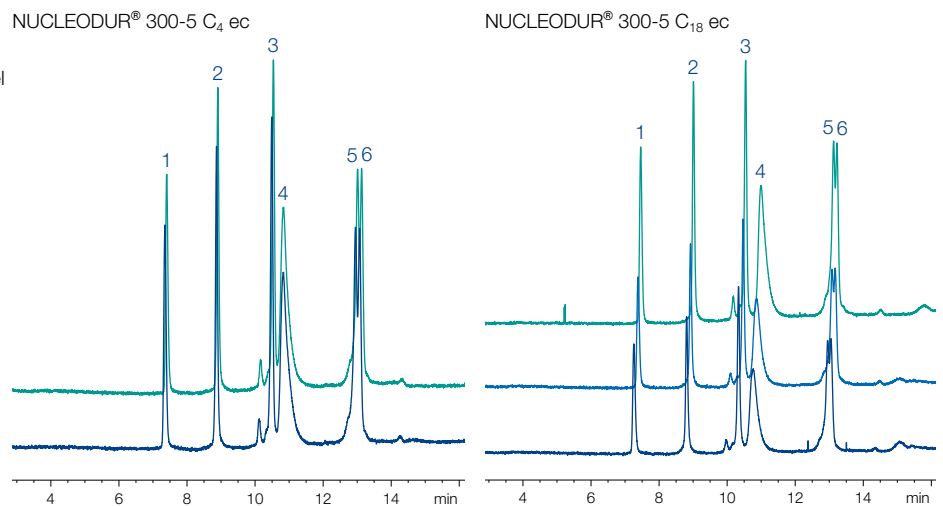


### Batch-to-batch reproducibility of NUCLEODUR® 300-5 C<sub>4</sub> ec and NUCLEODUR® 300-5 C<sub>18</sub> ec

MN Appl. Nos. 126551 / 126552

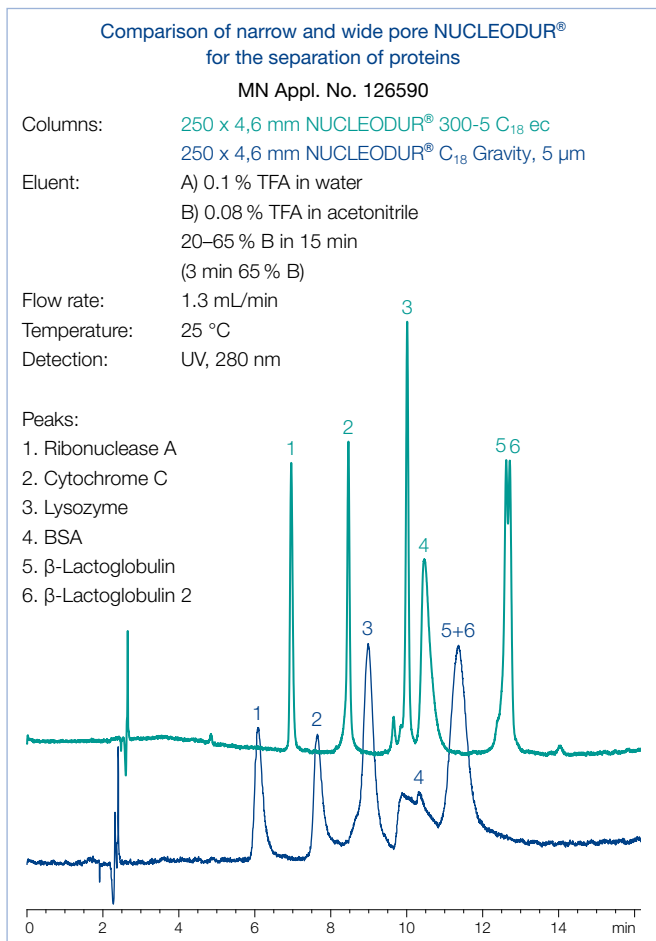
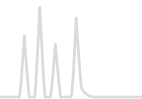
Columns: 250 x 4 mm  
 Eluent: A) 0.1 % TFA in water  
 B) 0.08 % TFA in acetonitrile  
 20–60 % B in 15 min  
 Flow rate: 1 mL/min  
 Temperature: 25 °C  
 Detection: UV, 280 nm

Peaks:  
 1. Ribonuclease A  
 2. Cytochrome C  
 3. Lysozyme  
 4. BSA  
 5. β-Lactoglobulin  
 6. β-Lactoglobulin 2

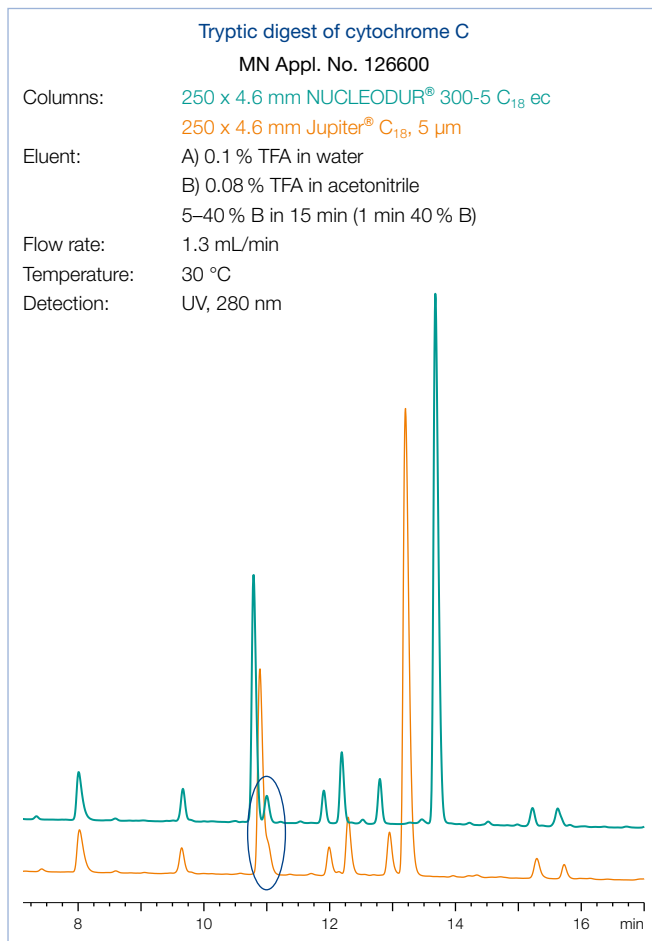




# HPLC columns for biochemical separations





Sharper peaks of larger molecules on wide pore material



Less tailing and better separation on NUCLEODUR® 300 C<sub>18</sub> ec

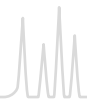
## Ordering information

Eluent in column acetonitrile – water

ID	Length →					EC guard columns*
	100 mm	125 mm	150 mm	250 mm		
<b>NUCLEODUR® 300-5 C<sub>18</sub> ec</b> octadecyl phase, particle size 5 μm, pore size 300 Å, endcapped, 4 % C						
Analytical EC columns						
	2 mm	760183.20	760184.20	760185.20	760186.20	761988.20
	3 mm	760183.30	760184.30	760185.30	760186.30	761988.30
	4 mm	760183.40	760184.40	760185.40	760186.40	761988.30
	4.6 mm	760183.46	760184.46	760185.46	760186.46	761988.30
<b>NUCLEODUR® 300-5 C<sub>4</sub> ec</b> butyl phase, particle size 5 μm, pore size 300 Å, endcapped, 2.5 % C						
Analytical EC columns						
	2 mm	760193.20	760194.20	760195.20	760196.20	761989.20
	3 mm	760193.30	760194.30	760195.30	760196.30	761989.30
	4 mm	760193.40	760194.40	760195.40	760196.40	761989.30
	4.6 mm	760193.46	760194.46	760195.46	760196.46	761989.30

\* EC guard columns require the Column Protection System guard column holder (REF 718966, see page 251).

EC columns in packs of 1, guard columns in packs of 3.



## NUCLEOSIL® MPN RP chromatography of biological macromolecules

### NUCLEOSIL® 100-5 C<sub>18</sub> MPN · USP L1

#### ★ Key feature

- Octadecyl phase, particle size 5 µm; pore size 100 Å
- Dynamic protein binding capacity per g packing: 6 mg BSA, 110 mg cytochrome C
- pH working range 2–8, max. working pressure 250 bar

#### 🔧 Technical data

- Silica-based reversed phase materials with monomerically bonded alkyl chains, brush type structure predominantly hydrophobic forces with a small portion of hydrophilic interactions
- Maximum separation efficiency can be achieved when the injected protein mass does not exceed 1–2 % of the maximum protein loading capacity.

### NUCLEOSIL® 300-5 C<sub>4</sub> MPN · USP L26

#### ★ Key feature

- Butyl phase, particle size 5 µm, pore size 300 Å
- Dynamic protein binding capacity per g packing: 14 mg BSA, 27 mg cytochrome C especially suited for the purification of larger, hydrophobic peptides and very different proteins
- pH working range 2–8, max. working pressure 250 bar

#### 🔧 Technical data

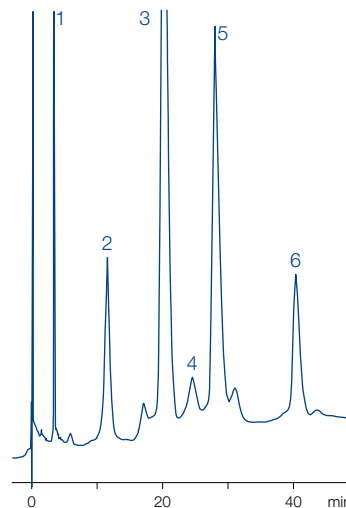
- Silica-based reversed phase materials with monomerically bonded alkyl chains, brush type structure predominantly hydrophobic forces with a small portion of hydrophilic interactions
- Maximum separation efficiency can be achieved when the injected protein mass does not exceed 1–2 % of the maximum protein loading capacity.

#### Separation of haemoglobin chains

MN Appl. No. 108240



Column: 250 x 4 mm NUCLEOSIL® 300-5 C<sub>4</sub> MPN  
 Eluent: A) 20 % acetonitrile, 80 % water, 0.1 % TFA  
 B) 60 % acetonitrile, 40 % water, 0.1 % TFA  
 40–60 % B in 60 min  
 Flow rate: 1 mL/min  
 Detection: UV, 220 nm

- Peaks:
1. Hem
  2. β-globin
  3. α-globin
  4. <sup>Δ</sup>γ<sup>T</sup>-globin
  5. <sup>ε</sup>γ-globin
  6. <sup>Δ</sup>γ<sup>L</sup>-globin

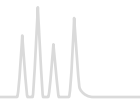


#### Ordering information

Eluent in column methanol

ID	Length → 250 mm	EC guard columns*
<b>NUCLEOSIL® 100-5 C<sub>18</sub> MPN</b>		
Analytical EC columns		
 4 mm	720231.40	
<b>NUCLEOSIL® 300-5 C<sub>4</sub> MPN</b>		
Analytical EC columns		
 4 mm	720245.40	721119.30

\* EC guard columns require the Column Protection System guard column holder (REF 718966, see page 251). Columns in packs of 1, guard columns in packs of 2.



## NUCLEOSIL® PPN RP chromatography of biological macromolecules

### NUCLEOSIL® 100-5 C<sub>18</sub> PPN · USP L1

#### ★ Key feature

- Octadecyl phase, particle size 5 µm, pore size 100 Å, dynamic protein binding capacity per g packing: 8 mg BSA, 64 mg cytochrome C; suited for the separation of peptides and proteins up to about 40 kD, also suited for basic peptides

#### 🔧 Technical data

- Silica-based reversed phase materials with polymerically bonded alkyl chains; exclusively hydrophobic interactions
- pH working range 1–9, max. working pressure 250 bar

### NUCLEOSIL® 500-5 C<sub>18</sub> PPN · USP L1

#### ★ Key feature

- Octadecyl phase, particle size 5 µm, pore size 500 Å, dynamic protein binding capacity per g packing: 22 mg BSA, 40 mg cytochrome C; especially suited for large peptides and medium-size hydrophilic proteins

#### 🔧 Technical data

- Silica-based reversed phase materials with polymerically bonded alkyl chains; exclusively hydrophobic interactions
- pH working range 1–9, max. working pressure 250 bar

#### Separation of a protein standard

MN Appl. No. 108220

Column: 125 x 4 mm NUCLEOSIL® 100-5 C<sub>18</sub> PPN

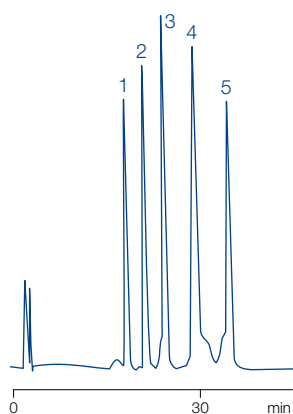
Eluent: A) 0.1 % TFA in H<sub>2</sub>O  
B) 0.08 % TFA in CH<sub>3</sub>CN  
20–60 % B in 10 min

Flow rate: 1.0 mL/min

Detection: UV, 280 nm

Peaks:

1. Ribonuclease
2. Cytochrome C
3. Lysozyme
4. β-Lactoglobulin
5. Ovalbumin



#### Separation of pancreatic secretion of piglets

MN Appl. No. 108280

Column: 125 x 4 mm NUCLEOSIL® 500-5 C<sub>18</sub> PPN

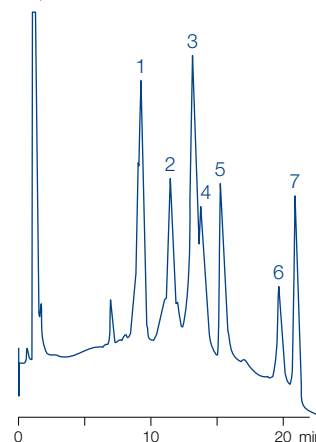
Eluent: A) 0.1 % TFA in H<sub>2</sub>O  
B) 0.08 % TFA in CH<sub>3</sub>CN  
30–50 % B in 14 min, then 50–65 % B in 6 min

Flow rate: 1 mL/min

Detection: UV, 215 nm

Peaks:

1. Trypsin + trypsinogen
2. Proelastase
3. Lipase + α-Chymotrypsin
4. Chymotrypsinogen
5. α-Amylase
- 6., 7. Procarboxypeptidase



## Ordering information

Eluent in column methanol

ID	Length → 250 mm	EC guard columns*
<b>NUCLEOSIL® 100-5 C<sub>18</sub> PPN</b> particle size 5 µm, pore size 100 Å		
Analytical EC columns		
4 mm	720252.40	721567.30
<b>NUCLEOSIL® 500-5 C<sub>18</sub> PPN</b> particle size 5 µm, pore size 500 Å		
Analytical EC columns		
4 mm	720258.40	721924.30

\* EC guard columns require the Column Protection System guard column holder (REF 718966, see page 251).

Columns in packs of 1, guard columns in packs of 2.



## NUCLEOGEL® RP columns RP columns for biochemical applications · USP L21

### Technical data

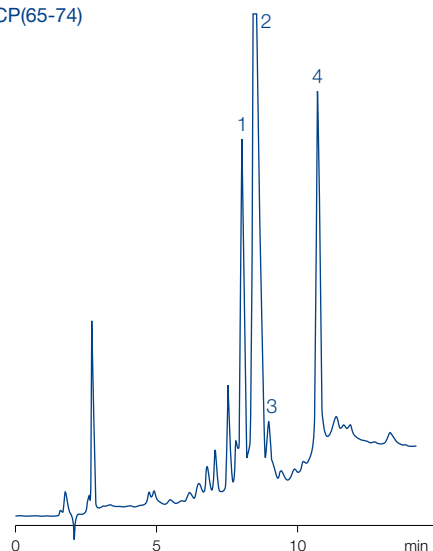
- Polystyrene resin cross-linked with divinylbenzene, available particle sizes 5 µm and 8 µm, available pore sizes 100 Å and 300 Å
- pH working range 1–13, max. working pressure 180 bar
- Small pore columns for reversed phase separation of small molecules such as pharmaceuticals with basic properties, e.g., organic heterocycles; also suited for separation of nucleosides and nucleotides up to 5000 Da; allow gradient as well as isocratic elution
- Wide pore columns are especially recommended for large biomolecules higher background hydrophobicity compared to silica phases

### Analysis of the synthetic acyl carrier protein ACP(65-74)

MN Appl. No. 108500

Column: 150 x 4.6 mm NUCLEOGEL® RP 100-8  
 Eluant: A) 0.1 % TFA in acetonitrile – water (1:99, v/v)  
 B) 0.1 % TFA in acetonitrile – water (99:1, v/v)  
 10–60 % B in 20 min  
 Flow rate: 1 mL/min  
 Detection: UV, 220 nm

- Peaks:
- ACP(66-74)(H-Gln)
  - ACP(65-74)
  - ACP(66-74)(Glp)
  - Thioanisole



### Ordering information

Eluent in column acetonitrile – water

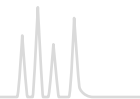
ID	Length →			Guard columns*
	50 mm	150 mm	250 mm	
<b>NUCLEOGEL® RP 100-5</b> particle size 5 µm, pore size 100 Å				
Analytical Valco type columns				
4.6 mm		719454	719455	719542
<b>NUCLEOGEL® RP 100-8</b> particle size 8 µm, pore size 100 Å				
Analytical Valco type columns				
4.6 mm		719456	719520	719542
<b>NUCLEOGEL® RP 300-5</b> particle size 5 µm, pore size 300 Å				
Analytical Valco type columns				
4.6 mm	719459			719542
<b>NUCLEOGEL® RP 300-8</b> particle size 8 µm, pore size 300 Å				
Analytical Valco type columns				
4.6 mm	719460			719542

\* Valco type guard columns measure 5 x 3 mm and require Guard column holder B, REF 719539, see page 250.

Columns in packs of 1, guard columns in packs of 2.



# HPLC columns for sugar analyses



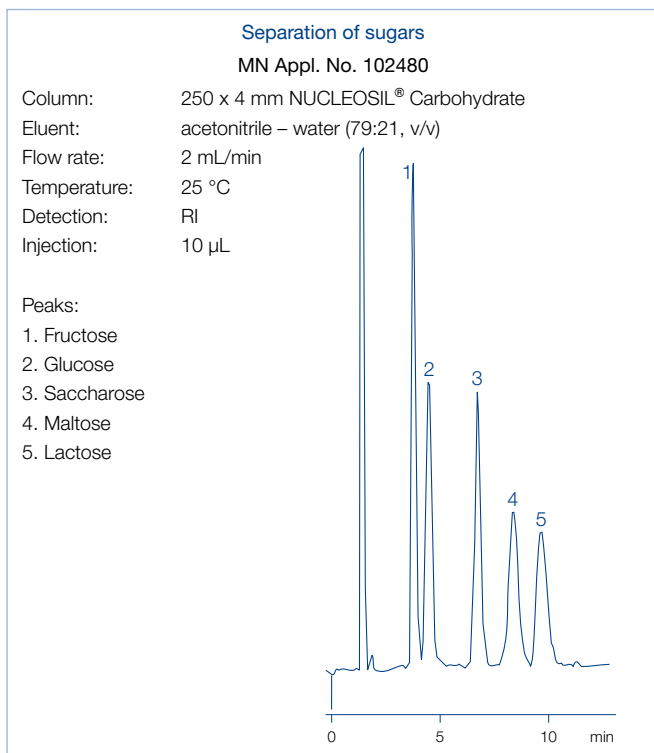
## NUCLEOSIL® Carbohydrate separation of mono- and disaccharides · USP L8

### Technical data

• Matrix: NUCLEOSIL® silica with amino modification, particle size 10 µm

### Recommended application

• RP separation of mono- and disaccharides



### Ordering information

Eluent in column acetonitrile – water (79:21, v/v)

ID	Length → 250 mm	EC guard columns*
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### NUCLEOSIL® Carbohydrate

Analytical EC columns

4 mm	720905.40	721170.30
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\* EC 4/3 guard columns for EC columns with 4 mm ID require the Column Protection System guard column holder (REF 718966, see page 251). Columns and guard columns in packs of 1.



## NUCLEOGEL® SUGAR 810 separation of sugars · USP L17 (H-Form) · USP L19 (Ca form)

### Technical data

- Sulfonated polystyrene - divinylbenzene resins in different ionic forms; due to a different selectivity pattern compared to NUCLEOGEL® SUGAR columns, the range of application is considerably enlarged
- Separation mechanism: ion exclusion, ion exchange, size exclusion, ligand exchange, NP and RP chromatography

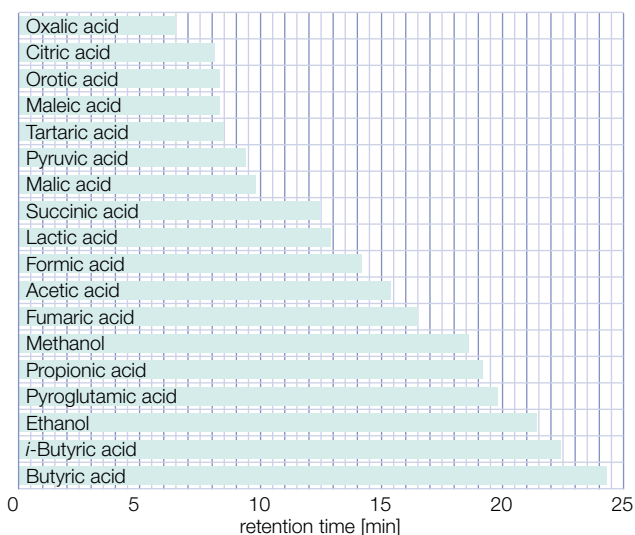
### Recommended application

- H<sup>+</sup> form: Separation of sugars, sugar alcohols and organic acids; eluent in column 5 mmol/L H<sub>2</sub>SO<sub>4</sub>
- Ca<sup>2+</sup> form: Separation of mono-, di- and oligosaccharides; eluent in column water

#### Organic acids and alcohols

MN Appl. No. 113870

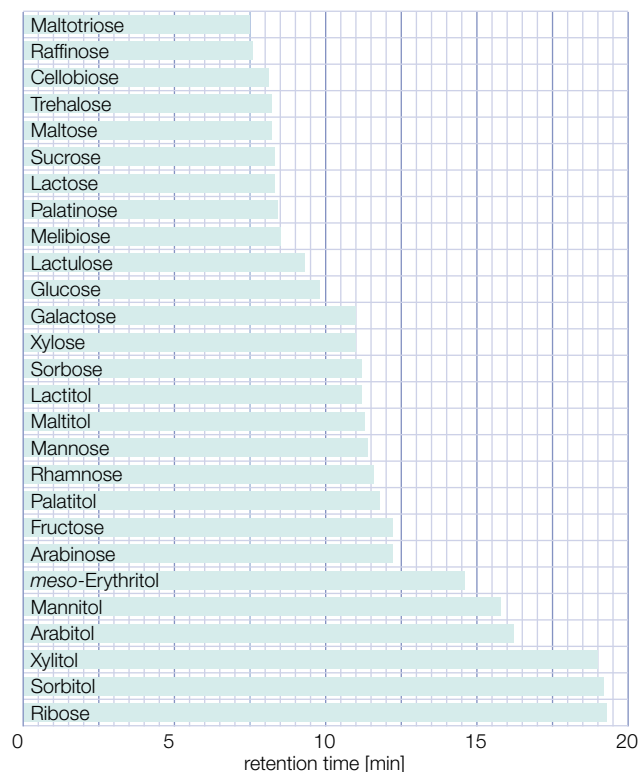
Column: 300 x 7.8 mm NUCLEOGEL® SUGAR 810 H  
 Eluent: 5 mmol/L H<sub>2</sub>SO<sub>4</sub>  
 Flow rate: 0.6 mL/min  
 Temperature: 35 °C  
 Detection: RI  
 Injection: 5 µL



#### Sugars and sugar alcohols

MN Appl. No. 114160

Column: 300 x 7.8 mm NUCLEOGEL® SUGAR 810 Ca  
 Eluent: water  
 Flow rate: 0.6 mL/min  
 Temperature: 85 °C  
 Detection: RI



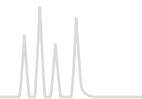
### Ordering information

ID	Length → 300 mm	Guard columns*
<b>NUCLEOGEL® SUGAR 810 H</b> eluent in column 5 mmol/L H <sub>2</sub> SO <sub>4</sub>		
Analytical Valco type columns		
7.8 mm	719574	719575
<b>NUCLEOGEL® SUGAR 810 Ca</b> eluent in column water		
Analytical Valco type columns		
7.8 mm	719570	719571

\* NUCLEOGEL® SUGAR 810 guard columns measure 30 x 4 mm and require the CC column holder 30 mm (REF 721823)  
 Columns in packs of 1, guard columns in packs of 2.



# HPLC columns for sugar analyses



## NUCLEOGEL® ION 300 OA / SUGAR

separation of sugars · USP L17 (H form) · USP L19 (Ca form) · USP L34 (Pb form) · USP L58 (Na form)

### Technical data

- Sulfonated spherical PS/DVB resins in different ionic forms; mean particle size 10 µm, pore size 100 Å
- Separation mechanism includes steric exclusion, ligand exchange and partition effects, ligand exchange being the predominant force, since the hydrated metal ions form strong interactions with the hydroxyl groups of the sample molecules. The intensity of these interactions decreases in the sequence Pb > Ca > Na
- Recommended operating temperatures: 60–95 °C; maximum pressure 70 bar

### Recommended application

- NUCLEOGEL® ION 300 OA:  
H<sup>+</sup> form for separation of sugars, alcohols and organic acids
- NUCLEOGEL® SUGAR:  
Ca<sup>2+</sup> form: separation of mono- and oligosaccharides, sugar alcohols
- Pb<sup>2+</sup> form: separation of mono- and disaccharides from food and biological samples
- Na<sup>+</sup> form: separation of oligosaccharides from starch hydrolysates and food

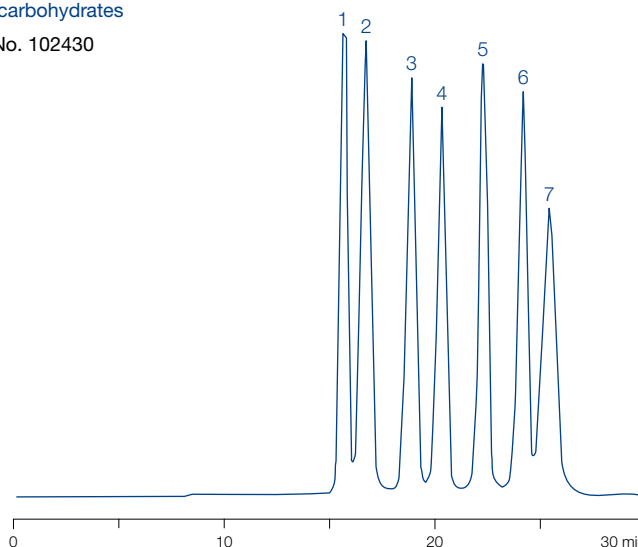
### Separation of carbohydrates

MN Appl. No. 102430

Column: 300 x 7.8 mm NUCLEOGEL® SUGAR Pb  
 Eluent: deionized water  
 Flow rate: 0.4 mL/min  
 Temperature: 80 °C  
 Detection: RI

#### Peaks:

1. Sucrose
2. Maltose
3. Glucose
4. Xylose
5. Galactose
6. Arabinose
7. Mannose



### Ordering information

ID	Length → 300 mm	Guard columns*
<b>NUCLEOGEL® ION 300 OA</b> eluent in column 5 mmol/L H <sub>2</sub> SO <sub>4</sub> 5 mmol/L H <sub>2</sub> SO <sub>4</sub>		
Analytical Valco type columns		
7.8 mm	719501	719537
<b>NUCLEOGEL® SUGAR Ca</b> eluent in column water + 0.02 % azide		
Analytical Valco type columns		
6.5 mm	719531	719535
<b>NUCLEOGEL® SUGAR Pb</b> eluent in column water + 0.02 % azide		
Analytical Valco type columns		
7.8 mm	719530	719534
<b>NUCLEOGEL® SUGAR Na</b> eluent in column water + 0.02 % azide		
Analytical Valco type columns		
7.8 mm	719532	719536

\* Valco Type guard columns measure 21 x 4 mm and require the guard column holder C, REF 719538, see page 250.  
 Columns in packs of 1, guard columns in packs of 2.

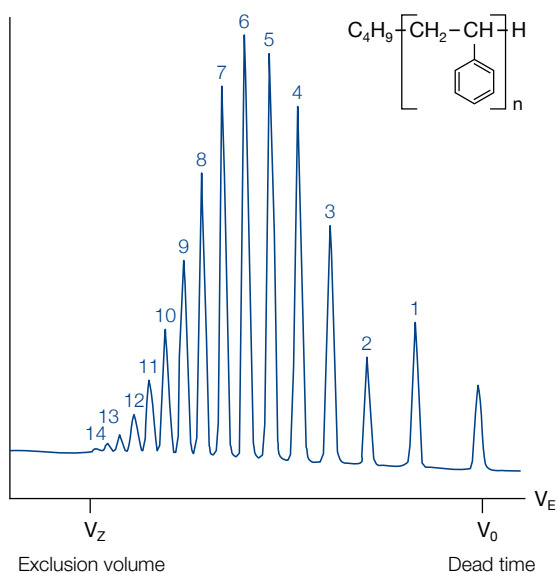


## NUCLEOGEL® GPC for GPC of water-insoluble substances

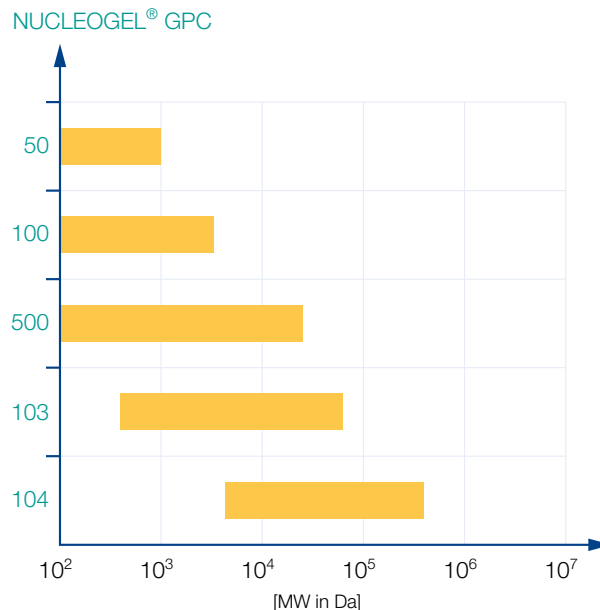
### Technical data

- Highly crosslinked macroporous, spherical polystyrene – divinylbenzene polymer matrix with good mechanical stability

### Chromatogram of styrene oligomers



### Working ranges for polystyrene



### Ordering information

Eluent in column toluene

Phase	Exclusion limit [kDalton]	Application	Column 300 x 7.7 mm	
<b>5 µm particle size</b>				
Analytical Valco type columns				
	NUCLEOGEL GPC 50	2	low molecular weight organics	719402
	NUCLEOGEL GPC 100	4	oligomers, oils	719403
	NUCLEOGEL GPC 500	25	low molecular weight polymers	719404
	NUCLEOGEL GPC 103	60	low molecular weight polymers	719405
	NUCLEOGEL GPC 104	500	polymers up to 500 kDa	719406
		guard columns 50 x 7.7 mm	719409	
<b>10 µm particle size</b>				
Analytical Valco type columns				
	NUCLEOGEL GPC 50	2	low molecular weight organics	719410
	NUCLEOGEL GPC 100	4	oligomers, oils	719411
	NUCLEOGEL GPC 500	25	low molecular weight polymers	719412
	NUCLEOGEL GPC 103	60	low molecular weight polymers	719413
	NUCLEOGEL GPC 104	500	polymers up to 500 kDa	719414
		guard columns 50 x 7.7 mm	719418	

Columns and guard columns in packs of 1.